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Abstract

This study assessed the impact of sonication on the structure and properties of a functional extract (retentate 100 or R100) from the Quorn fermentation co-product (centrate). In a previous study we reported that the R100 fraction displayed good foaming, emulsifying and rheological properties. Sonication of a R100 solution led to the breakdown of the large hyphal structures characteristic of this extract into smaller fragments. Foams prepared with sonicated R100 displayed a higher foaming ability than with untreated R100 and a high foam stability but lower than untreated R100 ones. Oil-in-water emulsions prepared with sonicated R100 displayed smaller oil droplet size distributions than with untreated R100. Confocal micrographs suggested that small fungal fragments contributed to the stabilisation of oil droplets. 50% oil-reduced R100 emulsions were prepared by mixing R100 emulsions (untreated or sonicated) with a sonicated R100 solution at a 1:1 ratio. Smaller oil droplet size distributions were reported for the oil-reduced emulsions. These results showed that the addition of small hyphal fragments or surface-active molecules and molecular aggregates released during sonication contributed to the formation and stabilisation of smaller oil droplets. This study highlighted the potential to modulate the structure, emulsifying and foaming properties of functional extracts from the Quorn fermentation co-product by sonication and the potential of these extracts as oil-lowering agents in emulsion-based products through the reduction of oil droplet size and their stabilisation.

Keywords

Quorn, co-product, centrate, sonication, emulsion, foam

Introduction

The World Health Organization (WHO) and the Food and Agriculture Organization of the United Nations (FAO) have recommended a reduction in daily intake of total fat to achieve a level of 15% to 30% of dietary energy [1-2]. As a

response academia and the food industry have been investigating a number of strategies to reduce the total fat content of food products while maintaining their structural and organoleptic properties. Of these, increasing the creaminess and satiety of emulsion-based formulations via the reduction of oil droplet size allows the possibility of reducing the oil content in products whilst maintaining hedonic and satiating properties. Oil-in-water emulsions (dispersions of oil into water as droplets stabilised by an emulsifier) are common food microstructures found in a wide range of products including sauces, condiments, spreads, dressings and desserts. The impact of oil droplet size on sensory perception and satiety has been explored in a series of nutritional intervention studies. A reduction in droplet size from 50 to 2 μm in a model oil-in-water drink formulation resulted in a significant 12.3% reduction in lunch intake among participants [3]. In another study, the creamy mouthfeel of a similar model drink formulation increased significantly as the oil droplet size was decreased from 50 to 11, 6, 2 and 0.2 μm while the hedonic appeal and fullness sensation increased and the hunger sensation recorded at 1 hour post-prandial decreased [4]. With regards to this strategy the food industry is investigating novel sustainable sources of functional ingredients and novel processes to increase their functionality with a view to reducing the oil droplet size in emulsion-based formulations.

In this context the production of mycoprotein by Marlow Foods for use in their meat alternative product Quorn is a potential source of sustainable functional ingredients. The term mycoprotein refers to the high-protein biomass produced by fermentation of the fungus *Fusarium venenatum* A3/5 (ATCC PTA-2684) by Marlow Foods, which forms the basis of their Quorn brand products. Mycoprotein contains all essential amino acids [5] and the net protein utilisation value of *Fusarium venenatum* mycoprotein is comparable to that of milk [6]. Moreover, the fermentation of starch into protein by *F. venenatum* results in 90% lower emission of greenhouse gases and benefits in terms of land and water footprints in comparison with beef products [7].

Following fermentation the broth undergoes an RNA-reduction step during which it is first heat-shocked above 68°C for 30 to 45 minutes, further heated at 90°C and finally centrifuged [8]. This heat shock step stops growth, disrupts ribosomes and activates endogenous RNAases which break down cellular RNA to nucleotides [8]. The resulting solid deposit (mycoprotein biomass) is then processed into a dough ready for conversion to the meat-like texture characteristic of Quorn foods. However the heat-shock RNA reduction step also induces diffusion of a fraction of the cell components through the cell wall [8]. The liquid supernatant (centrate) generated by the subsequent centrifugation step thus contains residual hyphal biomass, carbohydrates, nucleotides and proteins as well as the residues of the fermentation feedstock and is currently an unexploited co-product stream.

In a previous study we reported that an extract (retentate 100 or R100) obtained via a 100 kDa ultrafiltration of the centrate displayed good foaming stability, emulsifying and rheological properties (viscosity, viscoelasticity and gelation) [9]. R100 solutions displayed high viscosity, while heat-induced hydrogels of R100 solutions exhibited high viscoelasticity. R100 foams displayed high stability while oil-in-water R100 emulsions showed small and stable oil droplet size distributions. This study also reported that heat treatment (80°C for 30 mins) of a R100 solution increased its foaming ability, foam stability and emulsion stability. Large hyphal aggregates were reported in R100, contributing to its high viscosity and viscoelasticity and to foam and emulsion stabilisation. In addition tensiometry measurements showed that interfacially-active molecules present in R100 formed a rigid film at the oil/water interface. A number of functional metabolites and proteins that could have also contributed to the functional properties reported were

identified in the centrate. These included a cerato-platanin surfactant protein, cell membrane constituents (phospholipids, sterols, glycosphingolipids, sphingomyelins), cell wall constituents (chitin, chitosan, proteins) and guanine and guanine-based nucleosides and nucleotides.

Although centrate extracts have been shown to be functional, it is unknown whether this functionality can be improved through further processing. One of the novel processes used to modulate and/or increase the functionality of ingredients is the application of ultrasound through sonication. Ultrasound treatment has a wide range of applications in the food industry including homogenisation, extraction, reduction of particle size, cell destruction and dispersion of aggregates [10]. Of particular interest for this study is the reported breakdown of cell walls and the subsequent release of intracellular material into the medium as a result of the cavitation of bubbles and their implosion during sonication [11,12]. Sonication has more recently been applied to enhance the foaming and emulsifying properties of functional ingredients. The sonication of a range of proteins including egg white proteins, bovine serum albumin, soy proteins as well as a hydrophobin HFBII extract from the fungus *Trichoderma reesei* led to the stabilisation of air bubbles in solution, following which the subsequent mixing with an oil-in-water emulsion resulted in a triphasic air/oil/water system with potential for oil reduction [13-15].

The current study aimed to assess the possible modulation of the foaming and emulsifying properties of the R100 extract by sonication. In particular the impact of the application of ultrasound on the large hyphal structures present in R100 and the potential of sonicated R100 as emulsifier in an oil-reduced emulsion system were investigated.

Material and methods

Sample preparation

Centrate samples were collected from the Marlow Foods fermenter site at Belasis, Billingham (North Yorkshire) and frozen before analysis. Following thawing the centrate underwent a series of ultrafiltration steps using a range of Vivaflow 200 crossflow cassettes (Sartorius, UK) connected to a Masterflex Easy-Load peristaltic pump (Sartorius, UK). Following an initial 100 kDa ultrafiltration, the resulting retentate 100 or R100 (composed of material larger than 100 kDa) appeared as foamy, while the filtrate 100 underwent a further 50 kDa followed by a 5 kDa membrane, resulting in a retentate R100+50 (material between 50 and 100 kDa) and a retentate R100+50+5 (material between 5 and 50 kDa) fractions. Samples of the centrate and resulting retentates were freeze-dried in a Super Modulyo unit (Edwards, UK).

Nitrogen content

We previously reported that a wide range of functional molecules present in centrate and concentrated in the R100 fraction contained nitrogen, including fungal cell membrane and cell wall constituents such as phospholipids, glycosphingolipids, sphingomyelins, chitin, chitosan and proteins [9]. In that regard the nitrogen-containing material (NCM) content was measured using the Kjeldahl method (Lynch, Barbano, & Fleming, 1998) to provide a guideline for preparation of standardised sample quantities for functional tests [16]. Sets of 0.1 g of EW and R100 were digested in concentrated sulphuric acid (92%) using a Kjeltac Basic Digestion Unit 20 (Foss, UK) at 440°C in the presence of

a selenium catalyst. Distillation of the digested samples into boric acid was carried out using a Tecator Kjeltac 8100 Manual Distillation Unit (Foss, UK). The distilled samples were then titrated using 0.1 N hydrochloric acid. The % nitrogen was calculated using the following formula:

$$\% \text{ Nitrogen} = \frac{\text{Titration volume (ml)} \times 14.007}{\text{Sample weight (g)} \times 100}$$

The % nitrogen obtained was then multiplied by the general conversion factor of 6.25 recommended for mycoprotein biomass [17] to obtain a % nitrogen-containing material (NCM). Measurements were repeated three times.

Sonication

1% w/w NCM solutions of the centrate fractions were prepared in deionised water and stirred for one hour. The solutions were sonicated for 3 minutes at 20 kHz, 500 Watts and 50% amplitude using a Sonics Vibra-Cell VCX-500 probe sonicator (Sonics, UK).

Particle size distribution

The particle size distribution of the 1% w/w NCM untreated and sonicated centrate fraction solutions was assessed by laser light diffraction and dynamic light scattering. At the micrometre level the average particle size distribution (D[3,2] or surface weighted mean) was measured by laser light diffraction using a Beckman-Coulter LS 13 320 particle size analyser (Beckman-Coulter, UK). Since the refractive indexes (RIs) of the different untreated and sonicated centrate fractions were not known we used a method developed by Hayakawa, Nakahira, & Tsubaki (1995) [18] and Saveyn, Mermuys, Thas, & van der Meeren (2002) [19] to determine the optimal RI to use to derive particle size data for each sample. The particle size distributions of all samples were first measured using an RI of 1.40 and were then recalculated using RIs ranging from 1.35 to 1.80 with an interval of 0.05 via the associated LS 13 320 software (Beckman-Coulter, UK). The resulting D[3,2] values from these calculations were then plotted against the RI values for each sample. Based on the graphs obtained the optimal RI to use for each sample was determined (the criteria used in the determination of the optimal RI are provided in the results section). The laser obscuration was adjusted to 10% obscuration and triplicate measurements were carried out.

At the nanometre level the particle size distribution (radius) was measured by dynamic light scattering. The centrate fraction solutions were filtered using a 1µm cut-off membrane, placed in test-tubes and sealed. Measurements were performed using a 633 nm laser with scattered light collected at 90° from the incident beam direction. The data were processed using an ALV-LSE-5004 correlator and size distributions (radius) were obtained from mass-weighted correlation functions. Measurements were repeated three times.

Foaming properties

Two sets of 15 g solutions of the centrate protein fractions (1% w/w NCM) were prepared in 50 ml glass beakers, which corresponds to an initial volume of 18 cm³ (1.4 cm sample height and 4 cm beaker diameter), and stirred for one hour. One set of each solution was sonicated while the other set was left untreated. The solutions were frothed for 1 minute using a handheld whisk-type frother (Aerolatte, UK). The height of the resulting foam was measured

immediately after whisking and every 10 mins until collapse of the foam. The foaming ability was expressed as the initial height of the foam, while the foam stability was determined as the time needed for the foam to fully collapse. Measurements were conducted at room temperature and repeated twice.

Emulsifying properties

Two sets of 15 g solutions of the centrate fractions (standardised to 1% w/w NCM) were prepared in deionised water and stirred for one hour. One set of each solution was sonicated while the other set was left untreated. Each solution was then mixed with 5 g of sunflower oil (to obtain a 3:1 aqueous solution to oil phase ratio) and the mixture was homogenised using an IKA T18 Ultra-Turrax high-speed homogeniser (IKA-Werke GmbH, Germany) at 10,000 rpm for 1 min to produce an oil-in-water emulsion. The average oil droplet size distribution of the centrate fraction emulsions (D[3,2] or surface weighted mean) was measured by laser light diffraction with the refractive index set at 1.474 (corresponding to sunflower oil). The particle size distribution of the centrate fraction solutions (D[3,2]) was also measured using the same refractive index (1.474) in order to assess the potential influence of hyphal material on the oil droplet sizes and distributions obtained for emulsions prepared with centrate fractions.

Reduced-oil emulsions

The oil-lowering potential of the untreated and sonicated centrate fractions in emulsion-based systems was investigated. Pre-existing emulsion droplets and dispersions were combined as shown in Figure 1 in order to explore the contribution of the added emulsifying material to the interface and continuous phase. Two sets of 15 g solutions of each centrate fraction (1% w/w NCM) were stirred for one hour (Figure 1). One set of each solution was sonicated while the other set was left untreated (stirred). Oil-in-water emulsions were prepared with either the stirred or sonicated dispersions using the Ultra-Turrax homogeniser. The resulting emulsions were then mixed with either the stirred or sonicated dispersions at a 1:1 ratio using the high-speed homogeniser at 10,000 rpm for 1 min to produce a range of 15 g 50% oil-reduced emulsions. 50% oil emulsion controls were prepared with untreated and sonicated centrate fractions following the method described in the emulsifying properties section.

Confocal and bright-field microscopy

Untreated and sonicated centrate fraction solutions, foams and oil-in-water emulsions were imaged using a Leica TCS2 confocal laser scanning microscope (Leica Microsystems, Germany). Images were recorded at a resolution of 512 x 512 pixels and analysed using the associated DM SDK version 4.2.1 software. For the imaging of solutions and foams 1% w/w NCM solutions of the centrate fractions were supplemented with the fluorescent dye Rhodamine B. For hyphal staining in oil-in-water emulsions Rhodamine B was added to 1% w/w NCM centrate fraction solutions before mixing with the oil. The dye was excited at 514nm, the collection range was 600-700 nm and a 10x/0.25 dry lens was used. For oil droplet staining in oil-in-water emulsions traces of Nile Red dye (Sigma Aldrich Co., UK) were added to the oil prior to mixing with the centrate fraction solution. The dye was excited at 549 nm and fluoresced light was collected at the emission maximum of 628 nm. A 63x/1.40-0.60 oil lens was used. For the reduced-oil emulsifying experiments microscopic images of centrate fraction emulsions and oil-reduced emulsions were obtained using an

Olympus BX-50 light microscope equipped with a QImaging QICAM digital camera (Olympus, UK). Emulsions were observed using a 50x / 0.5 lens.

Results and discussion

Characterisation of untreated and sonicated solutions

The optimal refractive index (RI) to use to derive particle size distribution data for each centrate fraction sample (untreated and sonicated) was determined using a method developed by Hayakawa, Nakahira, & Tsubaki (1995) [18] and Saveyn, Mermuys, Thas, & van der Meeren (2002) [19]. Following the measurement of the particle size distributions (D[3,2]) of each sample using an RI of 1.40, the D[3,2] values were recalculated using RIs ranging from 1.35 to 1.80 with an interval of 0.05 and plotted against RI values for each sample. According to this method [18-19], the RI corresponding to the first peak of the recalculated particle size is considered to be the optimum for determining the particle size distribution. The D[3,2] values obtained for all the samples increased with RI values up to a certain RI value and then remained constant or decreased (results not shown). The optimal RI values (corresponding to the D[3,2] peak) obtained for untreated centrate, R100 and R100+50+5 solutions were respectively 1.38, 1.39 and 1.42, while the optimal RIs reported for the sonicated centrate, R100 and R100+50+5 samples were respectively 1.40, 1.41 and 1.42 (results not shown).

The particle size distribution of the 1% w/w NCM untreated and sonicated centrate fraction solutions was then assessed by laser light diffraction at the optimal RI determined for each sample (Figure 2a). Large particles were reported for untreated R100 solutions (composed of material larger than 100 kDa), with the highest peak observed at 234 μm and particles reported up to 864 μm . The highest peak displayed by the untreated centrate sample solution also corresponded to large particles (133 μm). In contrast small particles were reported for untreated R100+50+5 solutions (composed of molecules between 5 and 50 kDa), with the highest peak measured at 5 μm . Following sonication the large particles reported for the untreated centrate and R100 solutions were broken down into smaller ones, with major peaks reported at 4 μm for both samples. The sonication process did not significantly alter the particle size distribution of R100+50+5, with the major peak remaining at 5 μm .

The confocal micrographs confirmed the results obtained by laser light diffraction. Fungal cell and small hyphal debris were observed in the centrate (micrographs not shown). Due to the complex composition of the centrate a succession of ultrafiltration steps was designed to separate large centrate components (including fungal cell and hyphal debris and large molecular aggregates) from small molecules and oligomeric aggregates in order to assess the functional contribution of each fraction, using the unfiltered centrate as reference. Much larger hyphal structures were observed in R100 (composed of material larger than 100 kDa) (Figure 3a), resulting from the aggregation of the hyphal debris present in the centrate during their retention on the 100 kDa ultrafiltration membrane. As previously postulated the aggregation process observed could have resulted from the pressure exerted on the ultrafiltration membrane-retained material and/or from the temperature increase occurring during ultrafiltration.⁹ Confocal micrographs of sonicated R100 solutions showed that these large hyphal aggregates were broken down into smaller fragments as a result of the

sonication process (Figures 3b and 3c), supporting the view that the large structures observed in R100 following ultrafiltration formed due to interaction between smaller hyphal fragments.

The successive ultrafiltration steps resulted in an increase in nitrogen-containing material (NCM): 47.9% (centrate), 54.0% (R100) and 70.4% (R100+50+5). The NCM contents obtained for these three streams proved similar to the ones measured in our previous study (47.3%, 56.7% and 71.6%) [9]. Following the 100 kDa ultrafiltration step the retention and aggregation of hyphal debris in R100 (observed in Figure 3a) concentrated nitrogen-containing cell wall and cell membrane constituents (including phospholipids, sterols, glycosphingolipids, sphingomyelins, chitin, chitosan and glycoproteins) and contributed to the NCM increase in this sample. Following freeze-drying the weight of sample obtained for the R100+50 sample (composed of molecules between 50 and 100 kDa) was very low and insufficient to test the functional profile of this fraction. The final filtrate obtained (filtrate 100+50+5) did not prove functional (results not shown), indicating that the molecules smaller than 5 kDa initially present in the untreated centrate did not contribute to its functionality.

At the micrometre level both untreated centrate and R100 solutions displayed a major peak at 237 nm, with an additional peak reported at 376 nm for R100 (Figure 4a). Such particle sizes corresponded to molecular aggregates including protein oligomers. In contrast the highest peak observed for the untreated R100+50+5 sample was reported at 1 nm, corresponding to small molecules including protein monomers (Figure 4b). This 1 nm peak was also observed in the untreated centrate solution. Following sonication major additional peaks were observed at 2 and 3 nm respectively for the centrate and R100 samples, while the 376 nm peak increased for both centrate and R100 samples and the 237 nm increased for the centrate. The appearance of the 2 and 3 nm peaks suggested that the sonication process resulted in the rupturing of cell walls and the release of intracellular molecules and/or cell wall and cell membrane components (including phospholipids, chitin, chitosan, protein monomers or oligomers and small nucleoside or nucleoside chains). The cell walls would have been left largely intact during the ultrafiltration process, explaining the absence of the 2 and 3 nm peaks in untreated centrate and R100. However the increase of the high particle size peaks (376 nm in both centrate and R100 and 237 nm in centrate) coupled with the decrease or disappearance of lower particle size peaks (1 to 5 nm) in both centrate and R100 following sonication also suggested the additional aggregation of a number of 1-5 nm molecular species (including protein monomers or oligomers initially present in the samples and/or released from cells during sonication) into the larger molecular components (376 and/or 237 nm) as a result of the sonication process. Aggregation of proteins including bovine serum albumin, myoglobin and lysozyme following sonication has previously been reported as a result of enhanced intermolecular interactions including disulphide bridges facilitated by the unfolding of the native state of these proteins caused by cavitation [20]. The sonication process did not significantly alter the particle size distribution of R100+50+5, with the major peak remaining at 1 nm.

Foaming properties of untreated and sonicated solutions

Untreated R100 foams displayed a lower foaming ability but higher foam stability than the untreated centrate (Figure 5) in accordance with our previous findings.⁹ The sonication process resulted in an increase in foaming ability for the centrate and R100 samples. The stability of the sonicated R100 foam proved high, however not as high as that of the

untreated R100 foam. Sonicated R100 displayed higher foaming ability and stability than a sonicated commercial whey protein concentrate (WPC) control and sonicated R100+50+5 (results not shown).

High densities of air bubbles were observed in untreated and sonicated centrate foams (Figures 6a and 6b) in agreement with the high foaming abilities reported for centrate solutions (Figure 5) and with previous results obtained on untreated centrate [9]. In contrast few air bubbles were reported for R100 foams, although sonicated R100 foams (Figure 6d) displayed a higher concentration of air bubbles than untreated R100 ones (Figure 6c), which correlated with the increase in foaming ability measured for R100 following sonication (Figure 5). The poor foaming ability reported for untreated R100 was in agreement with our previous findings [9].

Confocal micrographs also showed that air bubbles in sonicated R100 foams were trapped in a less dense network of hyphal fragments (Figure 6d) than in untreated R100 foams (Figure 6c) due to the partial breakdown of R100 hyphal aggregates during sonication as observed in Figures 3b and 3c. This in turn allowed a higher number of air bubbles to be entrapped thus explaining the higher foaming ability of R100 following sonication. The dense hyphal network observed in untreated R100 solutions (Figure 3a) was previously associated with high viscosity [9], which only allowed a limited number of air bubbles to be formed and transported within the liquid, leading to a poor foaming ability. Such a mechanism agreed with existing literature in which highly viscous fermentation media resulted in issues with gas–liquid mass transfer for the filamentous fungus *Aspergillus terreus* [21] and in which sonication of the fermentation broth of the fungus *Cordyceps sinensis* Cs-HK1 caused a dramatic reduction of viscosity [22]. On the other hand the denser, more viscous hyphal network observed in untreated R100 foams better prevented the destabilisation of the air bubbles formed than in sonicated R100 foams by limiting their breakdown, probably through reduced drainage in the plateau borders, leading to a higher foam stability as reported in Figure 5.

In addition, as highlighted in the laser light diffraction results (Figure 2a), the breakdown of the R100 hyphal aggregates during sonication further released surface-active molecules, molecular aggregates and cell debris which could also have contributed to the high foaming ability of sonicated R100. Similarly the rupturing of fungal cell walls in sonicated centrate and R100 solutions as suggested by the dynamic light scattering results (Figure 4a) further released surface-active molecules, contributing to the increase in both foaming ability and stability of the centrate. Our previous study highlighted the presence of surface-active material in centrate fractions [9]. A number of proteins with known foaming properties were identified in centrate and R100. A cerato-platanin was concentrated in the R100 fraction in comparison with the centrate and the serpin SERPB1 was detected in both samples. The cerato-platanin EPL1 from the fungus *Trichoderma atroviride* was previously reported to form ordered self-assembled layers at the air/water interface [23] and hydrophobic/hydrophilic interfaces [24] while serpins Z4 and Z7 from barley have been characterised as foam-positive proteins in beer [25-26]. A number of metabolites with known foaming properties were also previously identified in centrate [9], including cell wall and cell membrane constituents such as chitin and chitosan [27-28] and intracellular metabolites such as monoglycerides and diglycerides [29], glycosyldiglycerides [30], triterpenoid and steroidal saponins [31], rhamnolipids [32] and fatty amides [33].

Emulsifying properties of untreated and sonicated solutions

Untreated R100 emulsions displayed a smaller oil droplet size than the untreated centrate, R100+50+5 (Figure 7a) and WPC control ones (results not shown), which was in accordance with our previous findings [9]. Sonicated R100 emulsions showed a smaller oil droplet size (mean oil droplet size of 13.57 μm) than untreated R100 ones (18.61 μm), however the sonication process did not yield a reduction in droplet size for centrate and R100+50+5. The mean oil droplet size in sonicated R100 emulsions also proved smaller than in sonicated WPC ones (23.75 μm , results not shown). Confocal micrographs confirmed the smaller oil droplet size distribution observed in sonicated R100 emulsions (Figure 8b) in comparison with centrate and R100+50+5 (Figures 8a and 8c).

A ring of small hyphal fragments was visible around oil droplets in untreated R100 emulsions (Figures 9b1 and 9c1), suggesting a role for the hyphal fragments in the formation and stabilisation of the emulsions. Similar observations were reported for sonicated R100 emulsions, however, due to the high inherent turbidity of sonicated R100 emulsions the micrographs did not appear as clear as untreated R100 ones at high magnification and the latter were chosen for illustration purpose in Figure 9. The size of the small hyphal fragments (1 to 4 μm) observed around oil droplets in sonicated R100 emulsions matched with the largest particle size peak reported at 4 μm in sonicated R100 solutions (Figure 2a), while the wider size range of fragments (2 to 18 μm) observed around oil droplets in untreated R100 emulsions (Figures 9b1 and 9c1) was reported at low level in the untreated R100 sample (Figure 2a). The shapes and sizes of bacterial cells, viruses and spores fall within the range for Pickering stabilisation of biphasic dispersions including foams and emulsions [34]. This in turn suggests that, based on their hydrophobicity, hyphal fragments present in R100 and/or released during the sonication and emulsifying processes contributed to the formation and stabilisation of small oil droplets in untreated and sonicated R100 emulsions through a Pickering-type stabilisation. A partially hydrophobic moss spore (*Lycopodium clavatum*) has been shown to stabilise oil-in-water emulsions [35], while the ability of certain intact bacterial cells to stabilise oil-in-water and water-in-oil emulsions has also been reported [36]. In a particle-stabilised R100 emulsion system the smaller hyphal debris reported in sonicated R100 in comparison with untreated R100 would appear to contribute to formation of smaller oil droplets than in untreated R100 emulsions, in agreement with the results presented in Figure 7a. Pickering stabilisation by natural biological particles has previously been reported, with bacterial cellulose nanocrystals [37] and cocoa particles [38] reported to form and stabilise oil-in-water emulsions. Pickering stabilisation by hyphal fragments could also explain the high foaming properties of untreated and sonicated R100 reported in Figure 5. A correlation between foam stability and cell surface hydrophobicity has previously been reported for different strains of *Acinetrobacter calcoaceticus* [39]. Moreover the hyphal network observed in untreated and sonicated R100 appeared to be entrapping oil droplets (Figure 9a1), which was in agreement with the high emulsion stability reported for untreated R100 in our previous study [9]. In addition to the release of hyphal fragments from R100 aggregates during sonication and emulsification, the rupturing of cell walls as suggested in Figure 4a also released molecules and molecular aggregates with known emulsifying properties, further contributing to the formation and stabilisation of small oil droplets. Emulsion-positive compounds previously reported in the centrate [9] included cell wall and cell membrane constituents (such as chitin and chitosan [27-28] and phospholipids [40]) and intracellular molecules (such as monoglycerides and diglycerides [29], glycosyldiglycerides [30], triterpenoid and steroidal saponins [31], rhamnolipids [32] and fatty amides [33]).

In parallel, emulsions were prepared with untreated and sonicated R100 solutions that were filtered using a 200nm cut-off membrane prior to emulsification. These solutions produced larger oil droplet size distributions (respective mean oil droplet sizes of 54.70 and 33.57 μm) than their non-filtered counterparts (Figure 7b). These results highlighted that, in addition to the small hyphal fragments identified in Figures 9b1 and c1, the larger hyphal fragments observed in Figure 9a1 also contributed to the emulsifying properties of sonicated R100 possibly by physically separating and limiting the movement of oil droplets.

The untreated R100+50+5 solution and all the sonicated centrate fraction solutions (centrate, R100 and R100+50+5) displayed a major particle size peak at 4 μm when measured by laser light diffraction at a refractive index of 1.474 corresponding to sunflower oil (Figure 2b). A major oil droplet size peak was reported at 8 μm for sonicated R100 emulsions (Figure 7), however this peak was absent for all other emulsion samples, showing that the presence of hyphal material in emulsions prepared with centrate fractions did not influence the measurement of their oil droplet sizes and distributions by laser light diffraction.

Characterisation of the reduced-oil emulsions

The mixing of sonicated R100 dispersion with either mixed or sonicated R100 emulsions (Figure 1) resulted in a decrease of their oil droplet size distributions: from 18.61 μm to 15.19 μm for the mixed R100 emulsion and from 13.57 μm to 8.50 μm for the sonicated R100 emulsion (Figure 10a). The 50% oil-reduced sonicated R100 emulsion displayed the lowest mean droplet size of all emulsions tested (8.50 μm). Similarly the mixed and sonicated oil-reduced centrate, R100+50+5 and WPC emulsions showed smaller oil droplet sizes than their full-oil counterparts (R100+50+5 and WPC results not shown), but not to the extent as the differences reported between oil-reduced and full-oil R100 emulsions. For oil-reduced centrate emulsions the oil droplet size diameter decreased from 27.70 μm to 17.80 μm for the mixed centrate emulsion and from 27.29 μm to 21.90 μm for the sonicated centrate emulsion. For all emulsions the R100 samples displayed the lowest oil droplet sizes. In contrast to the results obtained with the addition of sonicated dispersions to emulsions, the addition of stirred centrate, R100 or R100+50+5 dispersions to either mixed or sonicated emulsions did not reduce their oil droplet size (results not shown). The 50% oil emulsion controls prepared with untreated and sonicated centrate fractions displayed similar oil droplet size distributions to their full-oil counterparts (results not shown), showing that the decrease in oil droplet size reported for oil-reduced emulsions was due to the mixing with additional centrate fraction material rather than to the reduction in oil content. The light micrographs confirmed the smaller oil droplet size distributions reported for the 50% oil-reduced R100 emulsions (mixed emulsion / sonicated dispersion and sonicated emulsion / sonicated dispersion) (Figure 11). In particular the micrograph 11 of Figure 11 showed the very low droplet size distribution achieved when mixing a sonicated R100 emulsion with a sonicated R100 dispersion.

These results indicated that the addition of small hyphal fragments or other surface-active molecules present in sonicated R100 to the particle-stabilised R100 emulsion model observed in figure 9 resulted in the formation and stabilisation of smaller oil droplets. The presence of this material is likely to have contributed to the smaller droplet sizes by reducing the extent of droplet coalescence. The oil droplet size of the 50% oil-reduced R100 emulsions (mixed emulsion / sonicated dispersion and sonicated emulsion / sonicated dispersion) remained small and stable over a 7-

day period: from 15.19 μm to 16.56 μm for the mixed R100 emulsion and from 8.50 μm to 11.97 μm for the sonicated R100 emulsion (Figure 10b). The reduction of oil droplet size in an oil-reduced formulation has previously been reported to replicate the mouthfeel of a full-fat formulation [3-4], highlighting the potential of sonicated R100 as both emulsifier and oil-lowering agent in emulsion-based food products.

Conclusions

This study highlighted the potential to modulate the structure, emulsifying and foaming properties of a functional extract (retentate 100 or R100) from the Quorn fermentation co-product by sonication. Sonication of a R100 solution led to the breakdown of the large hyphal structures characteristic of untreated R100 into smaller fragments.

Foams prepared with sonicated R100 displayed a higher foaming ability than with untreated R100 as well as a high foam stability although lower than untreated R100 ones. The results indicated that the looser hyphal network observed in sonicated R100 foams and the associated release of small hyphal fragments and other surface-active molecules allowed for a higher number of air bubbles to be formed but did not stabilise them as well as the dense hyphal network observed in untreated R100 foams.

Oil-in-water emulsions prepared with sonicated R100 displayed smaller oil droplet size distributions than with untreated R100. Confocal micrographs of R100 emulsions suggested a Pickering-type stabilisation by small hyphal fragments released during sonication and emulsification. 50% oil-reduced R100 emulsions were prepared by mixing R100 emulsions with a sonicated R100 solution. This further addition of small hyphal fragments and other surface-active molecules resulted in even smaller oil droplet size distributions.

These results highlighted the potential of sonicated R100 as foaming agent, emulsifier and oil-lowering agent for the food industry. Additional studies will assess if the smaller oil droplet size measured in oil-reduced formulations allows to maintain the same creaminess and satiety sensation as full-fat formulations. Future studies will also be carried out to further understand the contribution of the hyphal fragments to the formation and stabilisation of emulsions and foams and to determine their optimal concentration and size for functionality. Finally this work highlights the possibility to investigate the impact of sonication on the mycelium biomass and functionality of the other Quorn fermentation process streams (fermentation broth and RNA-reduced broth).

Conflict of interest

There are no conflicts of interest to declare.

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Abbreviations

R100: retentate 100

NCM: nitrogen-containing material

WPC: whey protein concentrate

RI: refractive index

References

1. Food and Agriculture Organization of the United Nations (2010) Fats and fatty acids in human nutrition: report of an expert consultation. FAO Food and Nutrition 91.
2. World Health Organization (2003) Diet, Nutrition and the Prevention of Chronic Diseases. Technical Report No.916.
3. Lett AM, Norton JE, Yeomans MR (2003) Emulsion oil droplet size significantly affects satiety: A pre-ingestive approach. *Appetite* 96:18-24.
4. Lett AM, Yeomans MR, Norton IT, Norton JE (2016) Enhancing expected food intake behaviour, hedonics and sensory characteristics of oil-in-water emulsion systems through microstructural properties, oil droplet size and flavour. *Food Quality and Preference* 47(B):148-155.
5. Denny A, Aisbitt B, Lunn J (2008) Mycoprotein and health. *British Nutrition Bulletin* 33:298-310.
6. Edelman J, Fewell A, Solomons GL (1983) Myco-protein – a new food. *Nutr. Abstr. Rev. Clin. Nutr.* 53:471–480.
7. Finnigan TJA, Lemon M, Allen B, Patton I (2010) Mycoprotein LCA and Food 2030. *Aspects of Applied Biology* 102:81-90.
8. Ward PN (1996) A process for the reduction of nucleic acid content for a fungus imperfectus, WO Patent 95/23843.
9. Lonchamp J, Clegg PS, Euston SR (2019) Foaming, emulsifying and rheological properties of extracts from a co-product of the Quorn fermentation process. *European Food Research and Technology* 245(9):1825-1839.
10. Gallo G, Ferrara L, Naviglio D (2018) Application of Ultrasound in Food Science and Technology: A Perspective. *Foods* 7(164).
11. Feng H, Lee H (2011) Effect of Power Ultrasound on Food Quality. In: Feng H, Barbosa-Cánovas GV, Weiss J (ed) *Ultrasound Technologies for Food and Bioprocessing*. Springer, New York.
12. Clark JP (2008) An update on ultrasonics. *Food Technology* 26:75–77.
13. Tchuenbou-Magaia FL, Norton IT, Cox PW (2009) Hydrophobins stabilised air-filled emulsions for the food industry. *Food Hydrocolloids* 23:1877–1885.
14. Tchuenbou-Magaia FL, Al-Rifai N, Mohamed Ishak NE, Norton IT, Cox PW (2011) Suspensions of air cells with cysteine-rich protein coats: Air-filled emulsions. *Journal of Cellular Plastics* 47(3):217-232.
15. Suslick KS, Grinstaff MW, Kompany E, Wong M (1994) Characterization of sonochemically prepared proteinaceous microspheres. *Ultrasonics Sonochemistry* 1:S65–S68.
16. Lynch JM, Barbano DM, Fleming JR (1998) Indirect and direct determination of the casein content of milk by Kjeldahl nitrogen analysis: collaborative study. *Journal of AOAC International* 81:763-774.

17. Sadler MJ (2003) Mycoprotein. In: Caballero B, Trugo M, Finglas PM (ed) Encyclopedia of Food Sciences and Nutrition (Second Edition). Academic Press, Cambridge.
18. Hayakawa O, Nakahira K, Tsubaki JI (1995) Estimation of the optimum refractive index by the laser diffraction and scattering method on the raw material of fine ceramics. *Advanced Powder Technology* 6:47-61.
19. Saveyn H, Mermuys D, Thas O, van der Meeren P (2002) Determination of the refractive index of water-dispersible granules for use in laser diffraction experiments. *Particle & Particle Systems Characterization* 19:426-432.
20. Stathopoulos PB, Scholz GA, Hwang YM, Rumfeldt JAO, Lepock JR, Meiering EM (2004) Sonication of proteins causes formation of aggregates that resemble amyloid. *Protein Science* 13(11):3017-3027.
21. Porcel EMR, Casas Lopez JL, Sanchez Perez JA, Fernandez Sevilla JM, Chisti Y (2005) Effects of pellet morphology on broth rheology in fermentations of *Aspergillus terreus*. *Biochemical Engineering Journal* 26:139-144.
22. Cheung YC, Liu XX, Wang WQ, Wu JY (2015) Ultrasonic disruption of fungal mycelia for efficient recovery of polysaccharide-protein complexes from viscous fermentation broth of a medicinal fungus. *Ultrasonics Sonochemistry* 22:243-248.
23. Frischmann A, Neudl S, Gaderer R, Bonazza K, Zach S, Gruber S, Spadiut O, Friedbacher G, Grothe H, Seidl-Seiboth V (2013) Self-assembly at air/water interfaces and carbohydrate binding properties of the small secreted protein EPL1 from the fungus *Trichoderma atroviride*. *The Journal of Biological Chemistry* 288:4278-4287.
24. Bonazza K, Gaderer R, Neudl S, Przylucka A, Allmaier G, Druzhinina IS, Grothe H, Friedbacher G, Seidl-Seiboth V (2015) The fungal cerato-platanin protein EPL1 forms highly ordered layers at hydrophobic/hydrophilic interfaces. *Soft Matter* 11(9):1723-1732.
25. Li X, Jin Z, Gao F, Lu J, Cai G, Dong J, Yu J, Yang M (2014) Characterization of barley serpin Z7 that plays multiple roles in malt and beer. *Journal of Agricultural and Food Chemistry* 62(24):5643-50.
26. Specker C, Niessen L, Vogel RF (2014) In vitro studies on the main beer protein Z4 of *Hordeum vulgare* concerning heat stability, protease inhibition and gushing. *Journal of the Institute of Brewing* 120:85-92.
27. Lapasin R, Stefancic S, Delben F (1996) Rheological properties of emulsions containing soluble chitosan. *Agro-food Industry High-Tech* 7:12-17.
28. Quintela S, Villarán MC, López De Armentia I, Elejalde E (2012) Ochratoxin a removal from red wine by several oenological fining agents: Bentonite, egg albumin, allergen-free adsorbents, chitin and chitosan. *Food Additives & Contaminants: Part A* 29(7):1168-1174.
29. Moonen H, Bas H (2004) Mono- and diglycerides. In: Whitehurst RJ (ed) *Emulsifiers in Food Technology*. Blackwell Publishing Ltd, Oxford.
30. Keller RCA, Orsel R, Hamer RJ (1997) Competitive adsorption behaviour of wheat flour components and emulsifiers at an air-water interface. *Journal of Cereal Science* 25:175-183.
31. Kharkwal H, Panthari P, Pant MK, Kharkwal H, Kharkwal AC, Joshi DD (2012) Foaming glycosides: a review. *IOSR Journal of Pharmacy* 2(5):23-28.
32. Al-Ahmad K (2015) The Definition, Preparation and Application of Rhamnolipids as Biosurfactants. *International Journal of Nutrition and Food Sciences* 4(6):613-623.

33. Xu W, Gu H, Zhu X, Zhong Y, Jiang L, Xu M, Song A, Hao J (2015) CO₂-controllable foaming and emulsification properties of the stearic acid soap systems. *Langmuir* 31(21):5758-5766.
34. Lam S, Velikov KP, and Velev OD (2014) Pickering stabilization of foams and emulsions with particles of biological origin. *Current Opinion in Colloid & Interface Science* 19(5):490–500.
35. Binks BP, Clint JH, Mackenzie G, Simcock C, Whitby CP (2005) Naturally Occurring Spore Particles at Planar Fluid Interfaces and in Emulsions. *Langmuir* 21(18):8161–8167.
36. Dorobantu LS, Yeung AKC, Foght JM, Gray MR (2004) Stabilization of Oil-Water Emulsions by Hydrophobic Bacteria. *Applied and Environmental Microbiology* 70(10):6333-6336.
37. Kalashnikova I, Bizot H, Cathala B, Capron I (2011) New Pickering Emulsions Stabilized by Bacterial Cellulose Nanocrystals. *Langmuir* 27:7471–7479.
38. Gould J, Vieira J, Wolf B (2013) Cocoa particles for food emulsion stabilisation. *Food and Function* 4:1369–1375.
39. Rosenberg M, Rosenberg E (1985) Bacterial adherence at the hydrocarbon-water interface. *Oil and Petrochemical Pollution* 2(3):155-162.
40. Pichot R, Watson RL, Norton IT (2013) Phospholipids at the Interface: Current Trends and Challenges. *International Journal of Molecular Science* 14:11767-11794.

Figure captions

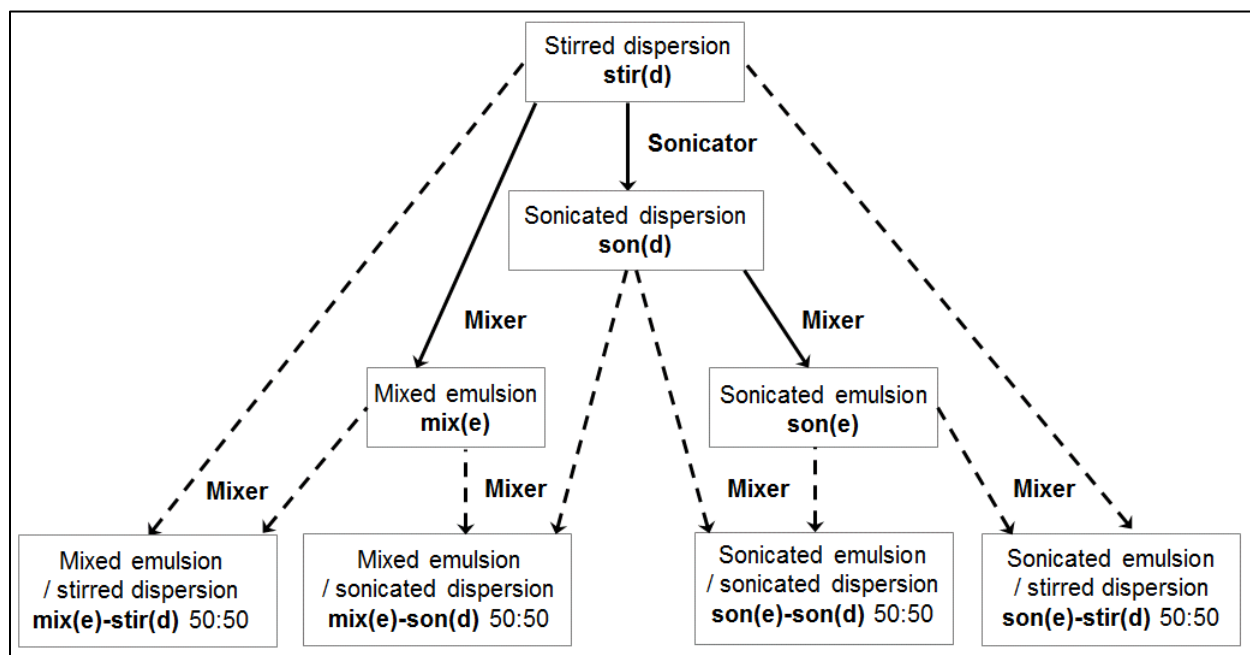


Fig. 1 Reduced-oil emulsion production diagram

d: dispersion, e: emulsion, stir: stirred, son: sonicated, mix: mixed

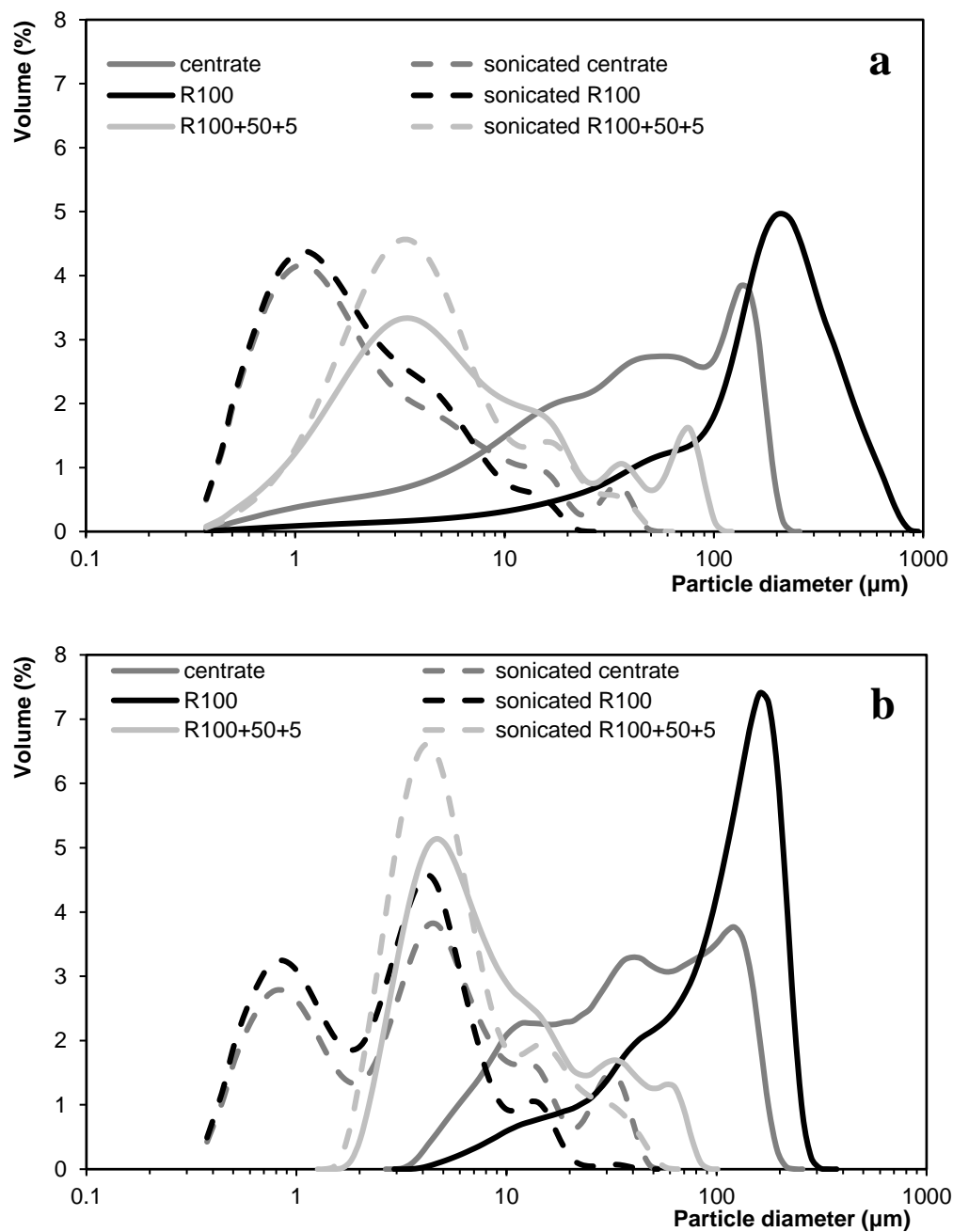


Fig. 2 Particle size distribution of untreated and sonicated centrate fraction solutions measured by laser light diffraction at the optimal refractive index determined for each sample (a) and at a refractive index of 1.474 (b) (1% w/w NCM, original pH, $n=3$)

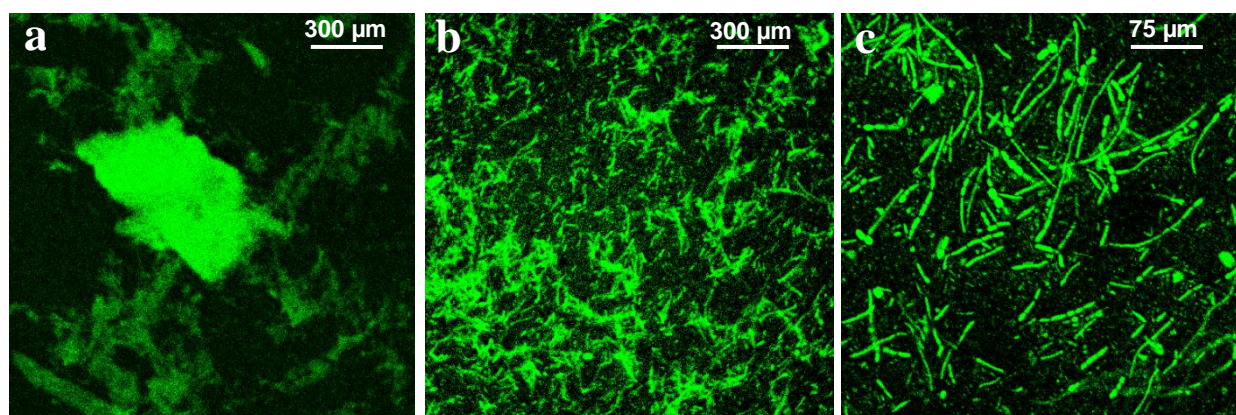


Fig. 3 Confocal microscopy of untreated and sonicated R100 solutions (1% w/w NCM, original pH, Rhodamine, magnification x10 [a,b] or x40 [c])

(a) R100, (b) sonicated R100, (c) sonicated R100

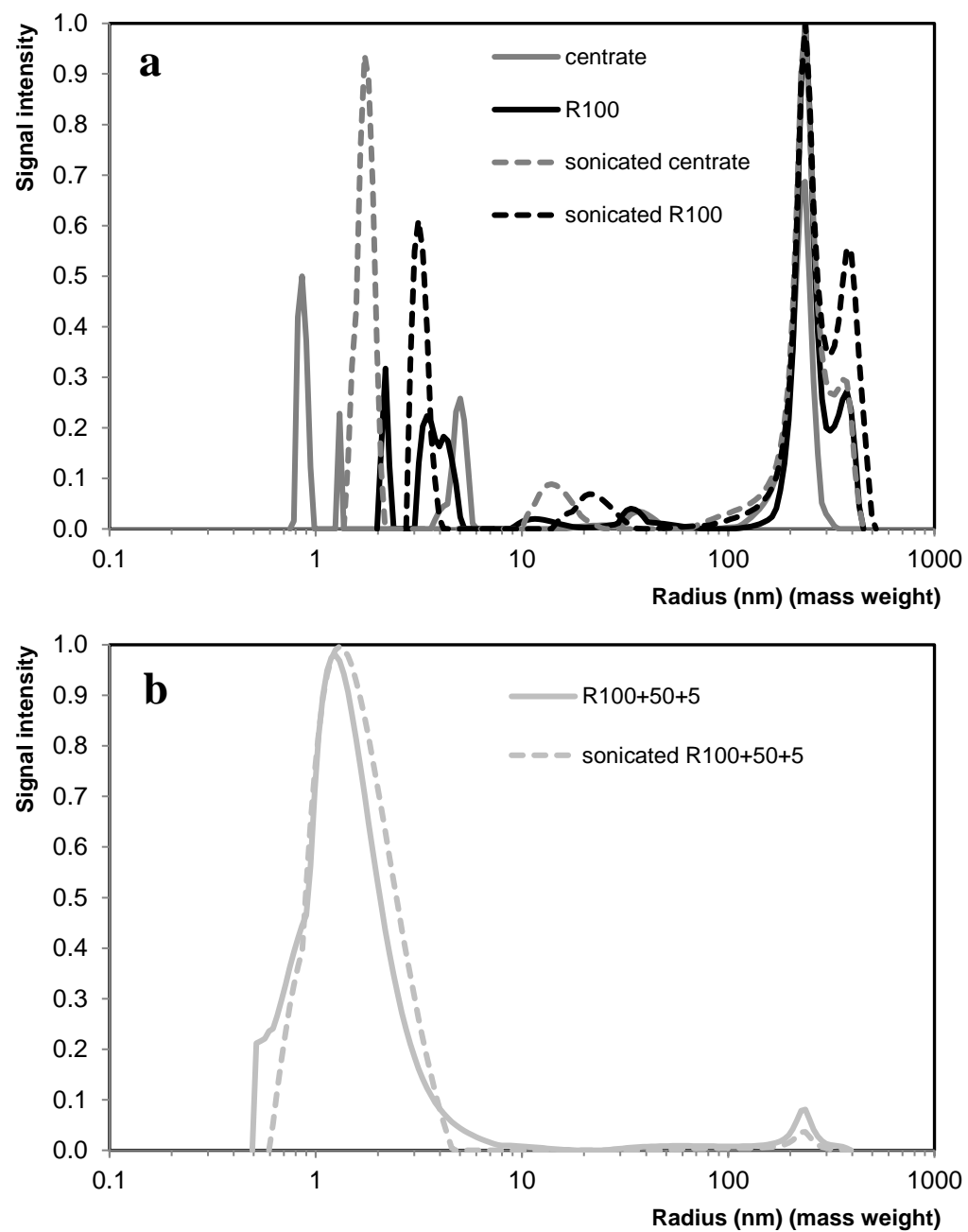


Fig. 4 Particle size distribution of untreated and sonicated centrate fraction solutions measured by dynamic light scattering (1% w/w NCM, original pH, $n=3$)

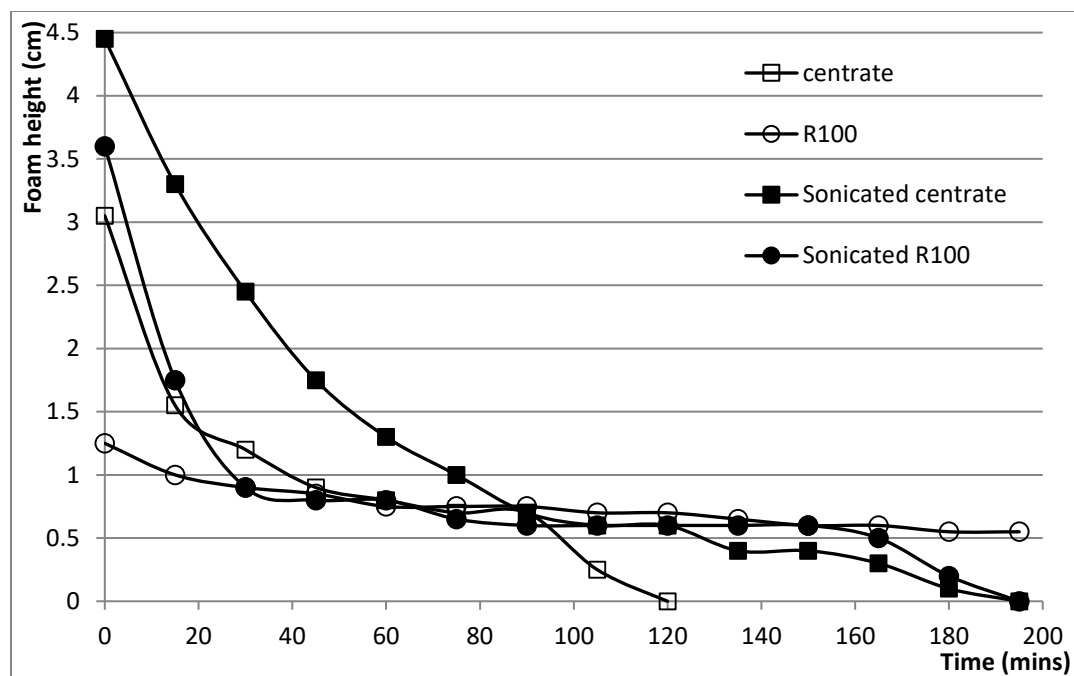


Fig. 5 Foaming ability and foam stability profiles of untreated and sonicated centrate fractions (frothing assay, 1% w/w NCM, original pH, n=3)

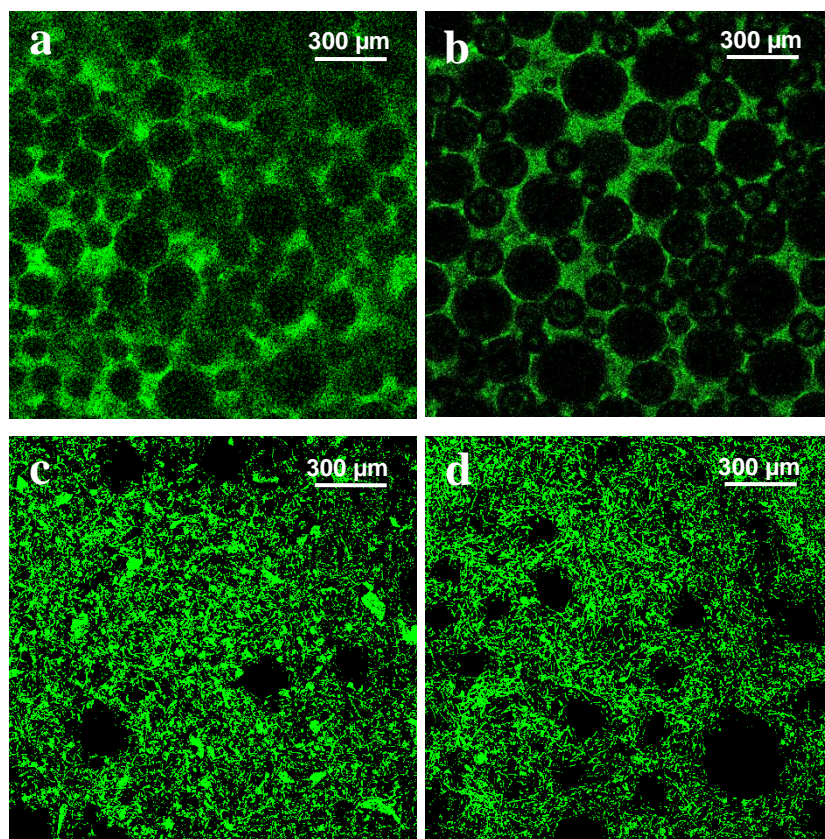


Fig. 6 Confocal microscopy of centrate fraction interfaces prepared by frothing (1% w/w NCM, original pH, Rhodamine, magnification x10)
(a) centrate foam, (b) sonicated centrate foam, (c) R100 foam, (d) sonicated R100 foam

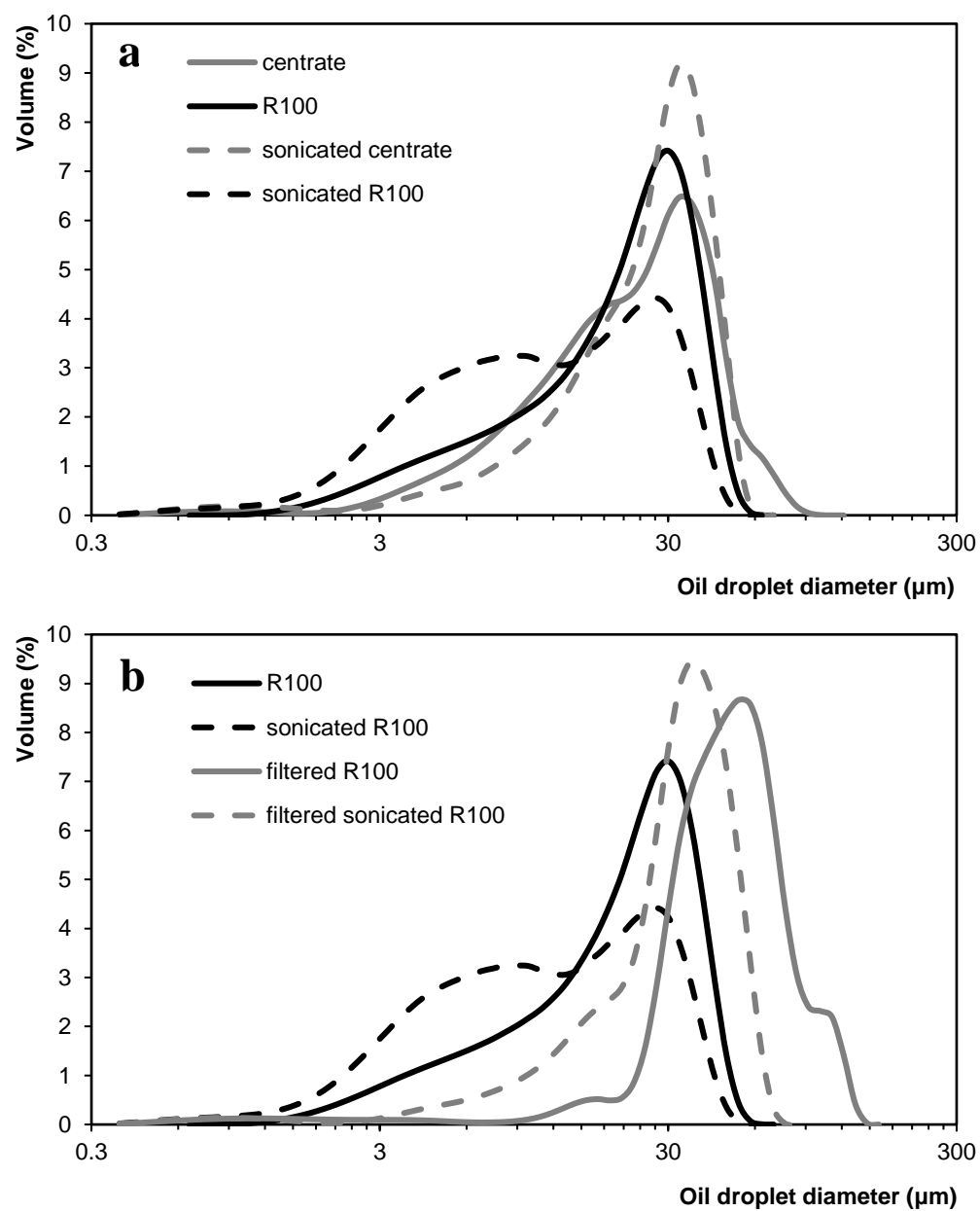


Fig. 7 Oil droplet size distribution of untreated and sonicated (a) and filtered untreated and sonicated centrate fraction emulsions (b) (1% w/w NCM, original pH, $n=3$)

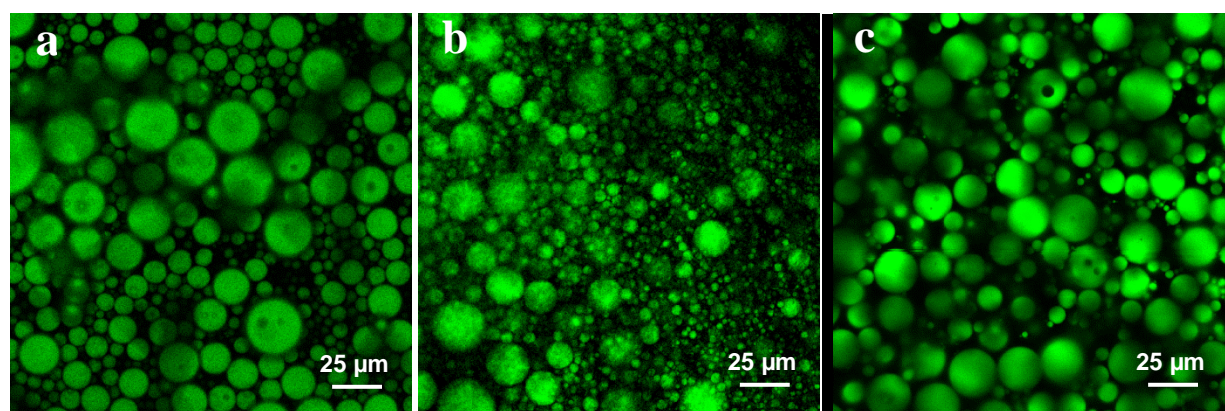


Fig. 8 Confocal microscopy of sonicated centrate fraction emulsions (1% w/w NCM, original pH, Nile Red, magnification x63)

(a) Centrate, (b) R100, (c) R100+50+5

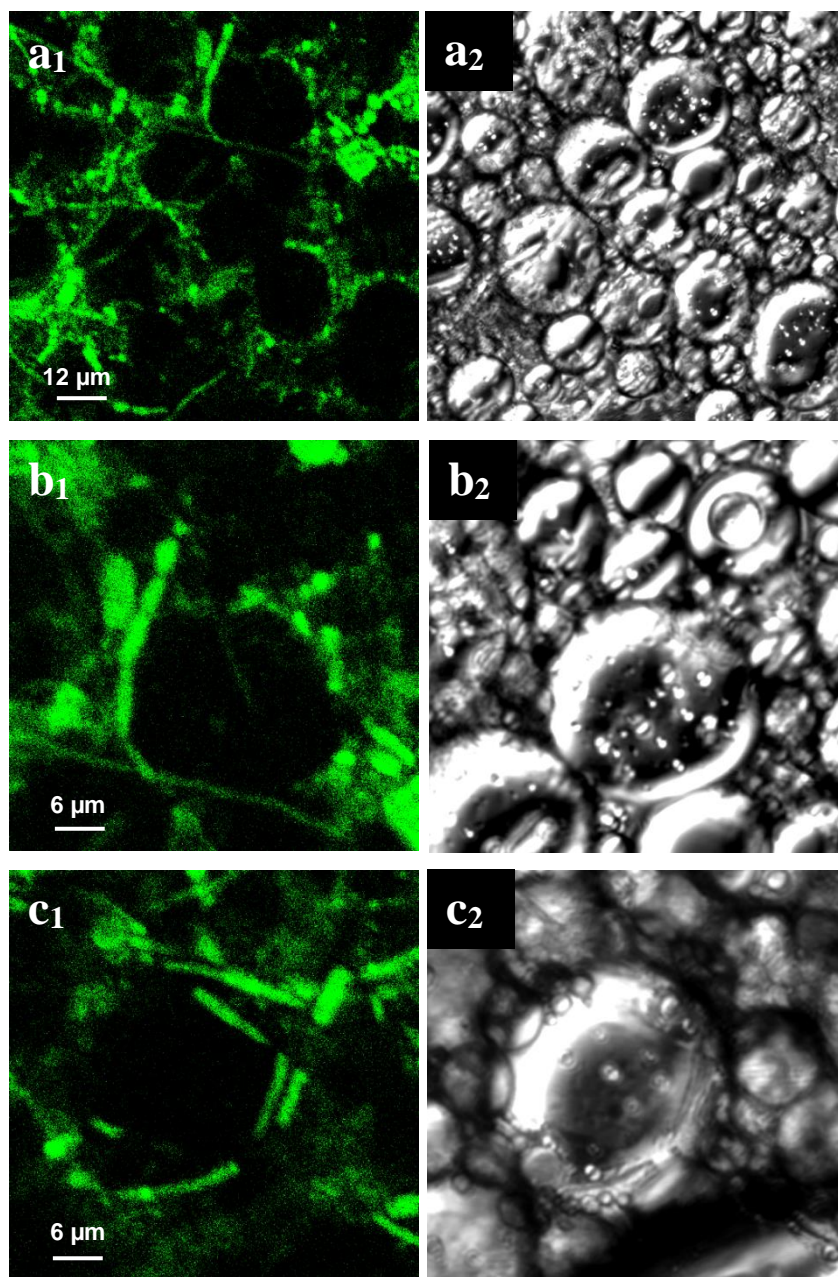


Fig. 9 Confocal microscopy of untreated R100 emulsions (1% w/w NCM, original pH, Rhodamine, magnification x63)

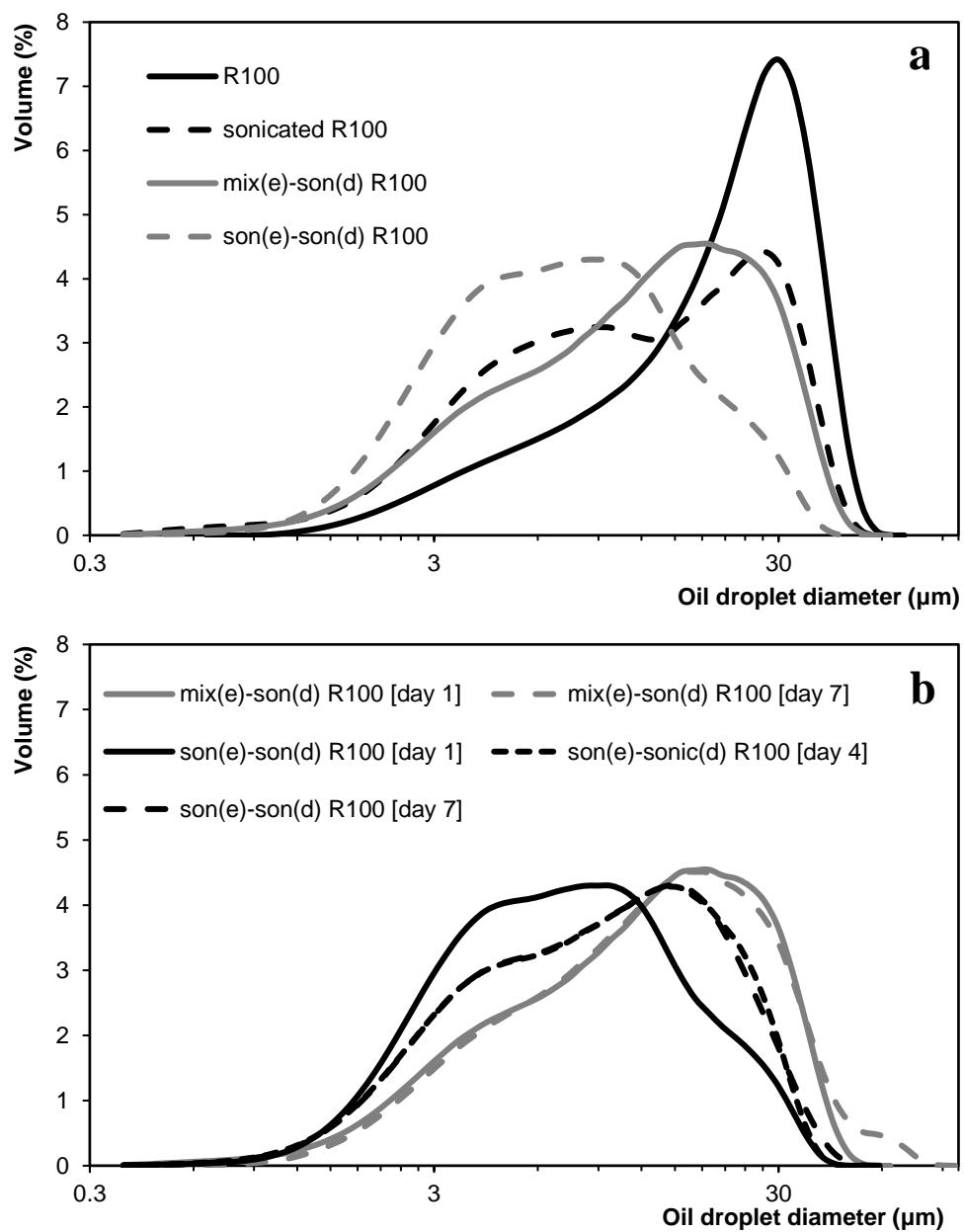


Fig. 10 Oil droplet size distribution of untreated, sonicated and oil-reduced R100 emulsions (a) and of oil-reduced R100 emulsions over 7 days of storage (b) (1% w/w NCM, original pH, n=3)

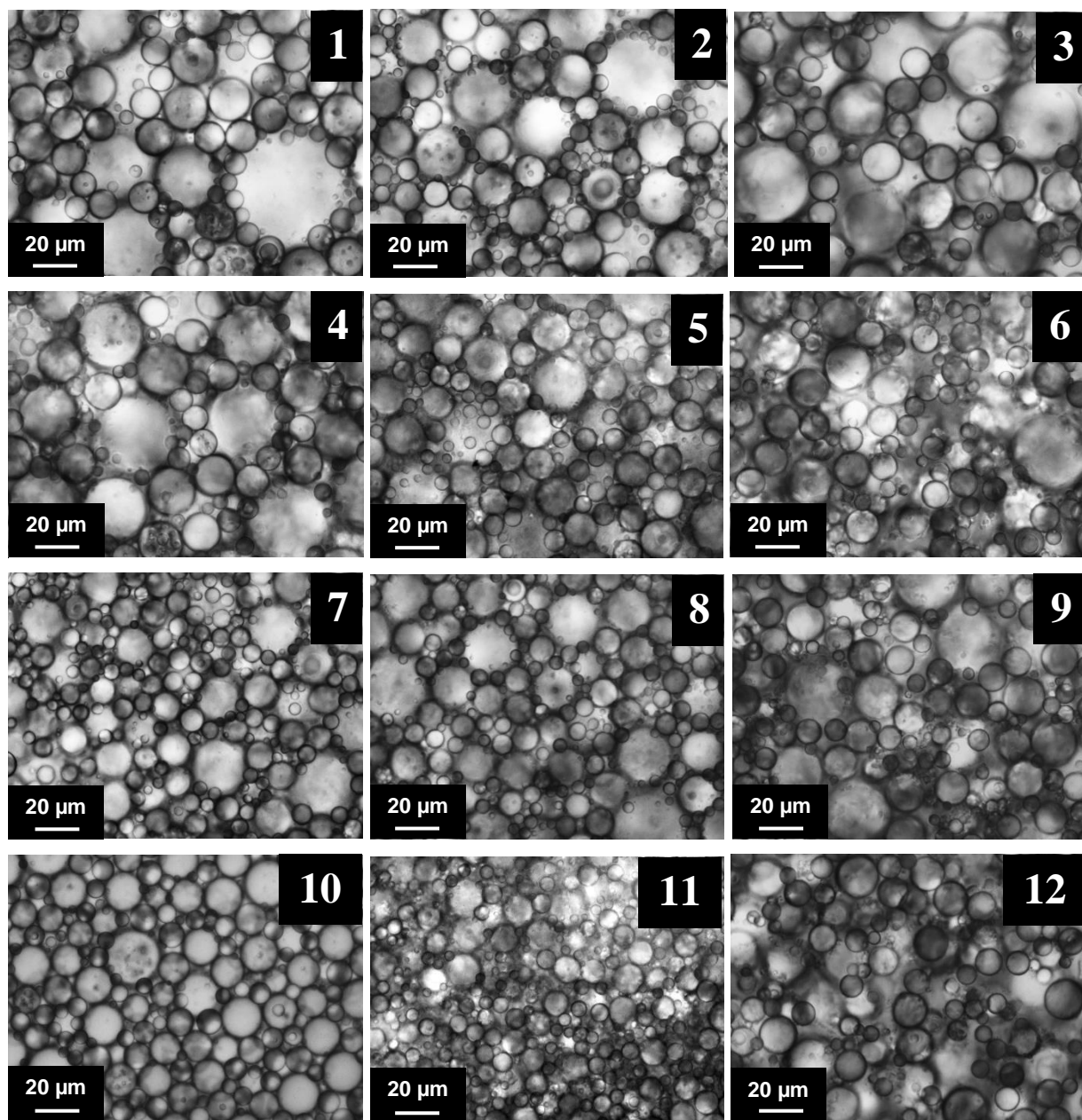


Fig. 11 Bright-field microscopy of full-oil and oil-reduced centrate fraction emulsions (1% w/w NCM, magnification x50)

(1) mix(e) centrate, (2) mix(e) R100, (3) mix(e) R00+50+5

(4) son(e) centrate, (5) son(e) R100, (6) son(e) R00+50+5

(7) mix(e)-son(d) centrate, (8) mix(e)-son(d) R100, (9) mix(e)-son(d) R00+50+5

(10) son(e)-son(d) centrate, (11) son(e)-son(d) R100, (12) son(e)-son(d) R00+50+5

e: emulsion, d: dispersion, mix: mixed, son: sonicated